Programmes for the eradication, control and monitoring of certain animal diseases and zoonoses

Survey programme for Avian Influenza (AI)

Approved* for 2012 by Commission Decision 2011/807/EU

Czech republic

* in accordance with Council Decision 2009/470/EC
1. Identification of the programme

Member state: CESKA REPUBLIKA

Disease: avian influenza in poultry and wild birds

Request of Community co-financing from beginning of: 2012 to end of 2012

1.1 Contact

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Fax: +420 227 010 195

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2. Description of the surveillance programme in poultry

2.1 Objectives of surveillance programmes

(max. 32000 chars):
The objectives of the surveillance programmes for avian influenza in poultry are to inform the competent authority of circulating avian influenza virus with a view to controlling the disease in accordance with Directive 2005/94/EC by the annual detection through active surveillance for:

(a) low pathogenic avian influenza (LPAI) of subtypes H5 and H7 in gallinaceous birds (namely chickens, turkeys, guinea fowl, pheasants, partridges and quails) and ratites thereby complementing other existing early detection systems;

(b) LPAI of subtypes H5 and H7 and highly pathogenic avian influenza (HPAI) in domestic waterfowl (namely ducks, geese and mallards for re-stocking supplies of game);

2.2 Design, implementation and target population
Standard requirements for the submission of surveillance programmes for avian influenza

version : 2.1

(max. 32000 chars) :

1. All positive findings will be retrospectively investigated at the holding and the conclusions of this investigation will be reported to the Commission and the CRL.

2. NRL will use specific protocols to accompany the sending of material to the CRL and reporting tables for collection of surveillance data. Protocols and tables are provided by the CRL. In those tables the laboratory testing methods used will be indicated.

3. Blood samples for serological examination will be collected from all species of poultry including those reared in free-range systems, 10 birds per holding (except ducks, geese and mallards -20 birds per holding), and from the different sheds, if more than one shed is present on a holding. In case of several sheds the sample size per holding will be increased appropriately. It is recommended to take at least 5 birds per shed 4. Sampling will be stratified throughout the territory of the whole Czech Republic. The stratification will be carry out by the State Veterinary Administration, taking into account:

(a) the number of holdings to be sampled (excluding ducks, geese and mallards); that number is defined so as to ensure the identification of at least one infected holding if the prevalence of infected holdings is at least 5%, with a 95% confidence interval ; and

(b) the number of birds sampled from each holding shall be defined so as to ensure 95% probability of identifying at least one positive bird if the prevalence of seropositive birds is more or equal 30%.

4. Based on a risk assessment and the specific situation the sampling design will also consider:

(a) The types of production and their specific risks, to be targeted to free range production, outdoor keeping plus taking into account other factors such as multi age, use of surface water, a relatively longer life span, the presence of more than one species on the holding or other relevant factors

(b) The number of duck, goose and mallards holdings to be sampled is defined to ensure the identification of at least one infected holding if the prevalence of infected holdings is at least 5%, with a 99% confidence interval.

(c) The time period for sampling will coincide with seasonal production. However, where appropriate, sampling will be adapted to other identified periods.

(d) Backyard flocks are not included into the surveillance programme.

2.2.1 Risk based surveillance (RBS)

(max. 32000 chars) :

The Czech Republic is out of the main migratory flight paths of birds from central and eastern Asia, the Caspian Sea and the Black sea areas. Due to this reason and according to the epidemiological and ornithological data we have not identified any region at higher risk for the introduction of AI. There are identified only areas with high density of poultry which is kept an open air holdings close to water areas
in the Czech Republic. For these reasons, the Czech Republic does not carry out this method (RBS) for surveillance for avian influenza.

### 2.2.2 Surveillance based on Representative Sampling

Surveillance based on a representative sampling scheme is implemented in the Czech Republic. Generally, the programme is applied on the whole territory of the Czech Republic, so that samples are considered as representative for the whole of the Czech Republic. The programme is implemented in all 14 regions in the Czech Republic. All regions are divided into the smaller areas which are named as districts. Programme is implemented on the regional level and Regional Veterinary Administrations (RVA) are in charge for performing of the programme.

### 3. Target populations

The surveillance programme for avian influenza in poultry will be included these categories:

- (a) laying hens;
- (b) free range laying hens;
- (c) turkey breeders;
- (d) duck breeders;
- (e) geese breeders;
- (f) fattening turkeys;
- (g) fattening ducks;
- (h) fattening geese;
- (i) farmed game birds (gallinaceous);
- (j) farmed game birds (waterfowl);

### 4. Risk-based surveillance (RBS) method

#### 4.1 Criteria and Risk factors

#### 4.1.1 Criteria and risk factors for virus introduction into poultry holdings due to direct or indirect exposure to wild birds in particular those of identified 'target species'
The Czech Republic does not carry out RBS method for surveillance for avian influenza. Surveillance based on a representative sampling scheme is implemented in the Czech Republic.

**4.1.2. Criteria and risk factors for virus spread within poultry holdings and between poultry holdings, as well as the consequences (impact) of the spread of avian influenza from poultry to poultry and between poultry holdings**

The Czech Republic does not carry out RBS method for surveillance for avian influenza. Surveillance based on a representative sampling scheme is implemented in the Czech Republic.

**4.2. Targeting of populations at risk**

The Czech Republic does not carry out RBS method for surveillance for avian influenza. Surveillance based on a representative sampling scheme is implemented in the Czech Republic.

**4.3. Targeting of poultry holdings to be sampled**

The Czech Republic does not carry out RBS method for surveillance for avian influenza. Surveillance
based on a representative sampling scheme is implemented in the Czech Republic.
### 5. Poultry holdings to be sampled

#### 5.1 Poultry holdings (except ducks, geese and mallard) to be sampled according to table 1 of Annex 1 to Decision 2010/367/EU

<table>
<thead>
<tr>
<th>NUTS (2) (a)</th>
<th>Total number of holdings</th>
<th>Total number of holdings to be sampled</th>
<th>Number of samples per holding</th>
<th>Total number of tests to be performed per method</th>
<th>Method of laboratory analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Czech Republic</td>
<td>126</td>
<td>53</td>
<td>10</td>
<td>530</td>
<td>ELISA test</td>
</tr>
</tbody>
</table>

Total

| Total        | 126                      | 53                                     | 10                           | 530                                           |                               |

(a)Refers to the location of the holding origin. In case NUTS 2 (Nomenclature of Territorial Units for Statistics) cannot be used, coordinates (longitude/latitude) are requested. Please fill-in these values directly in the field.

**Category:** laying hens

**Category:** free range laying hens
### Standard requirements for the submission of surveillance programmes for avian influenza

**version : 2.1**

<table>
<thead>
<tr>
<th>NUTS (2) (a)</th>
<th>Total number of holdings</th>
<th>Total number of holdings to be sampled</th>
<th>Number of samples per holding</th>
<th>Total number of tests to be performed per method</th>
<th>Method of laboratory analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>Czech Republic</td>
<td></td>
<td>5</td>
<td>5</td>
<td>10</td>
<td>50 ELISA test</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td></td>
<td><strong>5</strong></td>
<td><strong>5</strong></td>
<td><strong>10</strong></td>
<td></td>
</tr>
</tbody>
</table>

(a)Refers to the location of the holding origin. In case NUTS 2 (Nomenclature of Territorial Units for Statistics) cannot be used, coordinates (longitude/latitude) are requested. Please fill-in these values directly in the field.

#### Category: turkey breeders

<table>
<thead>
<tr>
<th>NUTS (2) (a)</th>
<th>Total number of holdings</th>
<th>Total number of holdings to be sampled</th>
<th>Number of samples per holding</th>
<th>Total number of tests to be performed per method</th>
<th>Method of laboratory analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>the Czech Republic</td>
<td></td>
<td>1</td>
<td>1</td>
<td>10</td>
<td>10 ELISA test</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td></td>
<td><strong>1</strong></td>
<td><strong>1</strong></td>
<td><strong>10</strong></td>
<td></td>
</tr>
</tbody>
</table>

(a)Refers to the location of the holding origin. In case NUTS 2 (Nomenclature of Territorial Units for Statistics) cannot be used, coordinates (longitude/latitude) are requested. Please fill-in these values directly in the field.
### Category: Fattening Turkeys

<table>
<thead>
<tr>
<th>NUTS (2) (a)</th>
<th>Total number of holdings</th>
<th>Total number of holdings to be sampled</th>
<th>Number of samples per holding</th>
<th>Total number of tests to be performed per method</th>
<th>Method of laboratory analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>the Czech Republic</td>
<td>72</td>
<td>42</td>
<td>10</td>
<td>420 ELISA test</td>
<td></td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td><strong>72</strong></td>
<td><strong>42</strong></td>
<td></td>
<td><strong>420</strong></td>
<td></td>
</tr>
</tbody>
</table>

(a) Refers to the location of the holding origin. In case NUTS 2 (Nomenclature of Territorial Units for Statistics) cannot be used, coordinates (longitude/latitude) are requested. Please fill-in these values directly in the field.

### Category: Farmed Game Birds (Gallinaceous)

<table>
<thead>
<tr>
<th>NUTS (2) (a)</th>
<th>Total number of holdings</th>
<th>Total number of holdings to be sampled</th>
<th>Number of samples per holding</th>
<th>Total number of tests to be performed per method</th>
<th>Method of laboratory analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>the Czech Republic</td>
<td>77</td>
<td>42</td>
<td>10</td>
<td>420 ELISA test</td>
<td></td>
</tr>
</tbody>
</table>

(a) Refers to the location of the holding origin. In case NUTS 2 (Nomenclature of Territorial Units for Statistics) cannot be used, coordinates (longitude/latitude) are requested. Please fill-in these values directly in the field.
### 5.2 Ducks, geese and mallard holdings to be sampled according to table 2 of Annex I to Decision 2010/367/EU

<table>
<thead>
<tr>
<th>NUTS (2) (a)</th>
<th>Total number of duck and goose holdings</th>
<th>Total number of duck and goose holdings to be sampled</th>
<th>Number of samples per holding</th>
<th>Total number of tests to be performed per method</th>
<th>Method of laboratory analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td>the Czech Republic</td>
<td></td>
<td>82</td>
<td>82</td>
<td>20</td>
<td>1,640 ELISA test</td>
</tr>
<tr>
<td>Total</td>
<td></td>
<td>82</td>
<td>82</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

(a) Refers to the location of the holding origin. In case NUTS 2 (Nomenclature of Territorial Units for Statistics) cannot be used, coordinates (longitude/latitude) are requested. Please fill in these values directly in the field.
6. Frequency and period for testing

(max. 32000 chars):

Birds in the poultry holdings will be sampled continuously during all year 2012. The time period for sampling in the poultry holding coincide with seasonal production for each poultry production category. Seasonal production is the most distinctive by fattening turkeys, ducks and geese, usually before Christmas and Easter. However, where appropriate, sampling will be adapted to other identified periods. The sampling of all poultry categories will be carried out once during 2012.

7. Laboratory testing

Description of the used serological tests: (max 4000 chars)

1. Laboratory tests will be carried out in accordance with the avian influenza diagnostic manual (Commission Decision 2006/437/EC) laying down the procedures for the confirmation and differential diagnosis of avian influenza.

2. All positive serological findings shall be confirmed by the NRL at SVI Prague for avian influenza by a haemagglutination-inhibition test, using designated strains supplied by the Community Reference Laboratory for Avian Influenza:
Standard requirements for the submission of
surveillance programmes for avian influenza
version : 2.1

H5
(a) Initial test using teal/England/7894/06 (H5N3)
(b) Test all positives with chicken/Scotland/59 (H5N1) to eliminate N3 cross reactive antibody.

H7
(a) Initial test using Turkey/England/647/77 (H7N7)
(b) Test all positives with African Starling/983/79 (H7N1) to eliminate N7 cross reactive antibody.

Testing of samples will be carried out in the National Reference Laboratory (NRL) in the State Veterinary Institute (SVI) in Prague and in SVIs in Olomouc and Jihlava. The NRL cooperates with Community Reference Laboratory for Avian Influenza (CRL) in Weybridge and the laboratories in SVIs Olomouc and Jihlava are under control of the NRL. All results to be sent to the Community Reference Laboratory for Avian Influenza (CRL) for collation.

8. Description of the surveillance programme in wild birds

8.1 Objectives of surveillance

Virological surveys for avian influenza in wild birds aim at identifying the risk for introduction of AI viruses (LPAI and HPAI) to domestic poultry by:

• ensuring of an early detection of HPAI H5N1 by investigating sick and dead wild birds and increased mortalities, in particular in selected “higher risk” species

• Anseriformes (wild fowl) to be the main sampling targets to assess if they carry LPAI viruses of H5 and H7 subtypes (which will in any case also detect HPAI H5N1 and other HPAI, if present)

• in case of HPAI H5N1 is detected in wild birds in an area, assessing specifically the risk for the introduction of HPAI H5N1 to poultry – by investigating dead and living wild birds of other bird species to possibly identify asymptomatic carriers and the risk that species coming in close contact with domestic...
poultry holdings might function as “bridge species”

- intensification of surveillance of HPAI H5N1 firstly in those areas where its presence has been confirmed

8.2 Surveillance design

(max. 32000 chars):

Passive surveillance on sick and dead wild birds to be targeted on:

- birds belonging to identified “higher risk” species (Mallard, Mute Swan, Greylag Goose, Comorant, Heron, Common Gull)
- areas where increased mortalities occur
- areas close to the ponds, lakes and waterways where birds were found dead; and in particular when these areas are in proximity to domestic poultry farms
- in areas where cases of HPAI H5N1 have been identified in wild birds or poultry to possibly identify asymptomatic carriers
- in areas epidemiologically linked to these cases

8.3 Sampling procedures

(max 32000 chars):

Taking samples, wrapping and packaging of samples and their transport to the NRL will be arranged by an official veterinarian from the relevant Regional Veterinary Administration.

- In the case of mortality increase (more than 5 birds) at least 5 carcasses to be taken. In the case of higher amounts of dead birds, cloacal and tracheal/oropharyngeal swabs form other birds can be taken as well.

- Cloacal swabs and tracheal/oropharyngeal swabs and/or tissues (namely the brain, heart, lung, kidney and intestines) from wild birds found dead or shot to be sampled for virus isolation and molecular detection (RT-PCR).
• Swabs must be chilled immediately on ice or with frozen gel packs and submitted to the laboratory as quickly as possible.

8.4 Laboratory testing

1. Laboratory tests to be carried out in accordance with the avian influenza diagnostic manual.

2. All samples collected in the survey for avian influenza in wild birds will be tested as soon as possible by molecular techniques and according to the diagnostic manual.

3. Initial screening using M gene PCR will be used, with rapid testing of positives for H5 (but within 2 weeks) and in case of a positive finding analysis of the cleavage site should be undertaken as soon as possible to determine whether or not it has a highly pathogenic avian influenza (HPAI) or a low pathogenic avian influenza (LPAI) motif.

4. All samples positive for M gene to be tested by virus isolation test on eggs.

5. At the laboratory pooling of up to five samples taken from the same species collected at the same site and same time is permitted when it can be ensured that, in case of a positive finding, the individual samples can be identified and retested.

6. Serological and virological results, as well as all isolates will be sent to the CRL.

8.5 WILD BIRDS - Investigation according to the surveillance programme for avian influenza in wild birds set out in Annex II to Decision 2010/367/EU
9. Description of the epidemiological situation of the disease in poultry during the last five years

max 4000 chars:

Over last year 2010 either highly pathogenic avian influenza or low pathogenic avian influenza subtype H5/H7 were not detected within the surveillance in poultry in the Czech Republic. There was only one positive finding of LPAI subtype H6N9 in the holding of farmed game birds (mallards) in poultry.

In 2009 there were detected 2 outbreaks of low pathogenic influenza in poultry holdings.

First outbreak: Hodonin
Low pathogenic avian influenza (LPAI) of subtype H7N9 was detected in poultry holding keeping goose and duck flocks in February 2009. The holding is situated on the ponds close to the city Hodonín, in Southern Moravian region, district Hodonin.
On pond were found infected geese. Samples were taken in the framework of routine AI surveillance in a poultry holding. There were 2 sheds of 679 heads of geese were killed using CO2 in gas-tight containers on the 25th February 2009.

Second outbreak: Třebín
LPAI of subtype H5N3 was detected in holding producing captive mallards for restocking in November 2009. The holding is situated on pond near municipality Třebín. There were kept 280 mallards for restocking. Samples were taken in the framework of routine AI surveillance in farmed game birds holding. All 280 mallards have been culled by private veterinarian used “T 61” on 6th November and safety disposed to the rendering plant.
In 2008 either highly pathogenic avian influenza (HPAI) or low pathogenic avian influenza (LPAI) were not detected in the Czech Republic.

In 2007 there was historically first outbreak of HPAI in poultry holding. Totally 4 holdings owned by two different farmers were affected in 2007. All holdings were situated in Pardubice Region, district Ústí nad Orlicí. Due to animal health reasons, 103 378 head of poultry (4 586 turkeys and a 98 792 head of species Gallus gallus) of total weight of 358 262 kg were killed in commercial flocks and another two contact holdings with the total number of poultry 67 879 head (54 125 broilers and 13 754 turkeys) were depopulated preventively. Furthermore, it was decided to kill preventively poultry kept in non-commercial flocks in municipalities situated in catastral territories of HPAI outbreaks (2 026 head of poultry).

The first outbreak was confirmed in a turkey flock in Tisová on 21 June 2007, the second outbreak was confirmed in a broiler flock in Nořín on 27 June 2007, the third outbreak was confirmed in a reproduction poultry flock of species Gallus gallus in Netřeby and the fourth outbreak was confirmed in a reproduction poultry flock of species Gallus gallus in Chocen, both outbreaks on the same day, i.e. on 11 July 2007.

Measures ordered in zones were repealed on 14 August 2007, in accordance with the legislation in force.

One year before, in 2006 avian influenza was detected only in wild birds, not in poultry.

9.1 Measures included in the programme for surveillance in poultry

9.1.1 Designation of the central authority in charge of supervising and coordinating the departments responsible for implementing the programme

(max. 4000 chars):

The central authority competent for supervising and coordinating all activities in veterinary care is the State Veterinary Administration, which performs its powers at the whole territory of the Czech Republic (§ 47, Veterinary Act No 166/1999 Col. of Acts). SVA of the CR coordinates the activities of Regional Veterinary Administrations (RVA). The surveillance programme for AI is laid down on the base of § 48 point 1, and § 10, Veterinary Act No. 166/1999 as amended.
Ministry of Agriculture of the Czech Republic determines main strategies in a veterinary care and carries out their control as laid down in the Veterinary Act No. 166/1999 Article 44, Point 1a. The Ministry of Agriculture specifies obligatory preventive and diagnostics campaigns in accordance with the Veterinary Act, Article 44; Point 1d, based on the epidemiological situation. Related details are laid down in the “Methodology of Animal Health Controls and Prophylaxis” approved by the Ministry of Agriculture and issued in its Official Journal. According to the legislation (Veterinary Act 166/1999) the SVA CR has the legal power to supervise any action ordered by the “Methodology”. Regional veterinary administrations execute the legal powers as to supervise private veterinarians over their actions in the professional field as ordered by the “Methodology”.

The competent authority confirms its commitment to submit a complete report on actions and expenditures in the framework of this programme and to provide additional information when they are request by the Commission.

9.1.2 System in place for the registration of holdings

Holdings in the Czech Republic are registered in the Database of Farms in accordance to the provisions of the law No. 154/2000, Breeding Act and corresponding Decree No.136/2004 laying down details for identification of animals and their registration and registration of holdings and person established by Breeding Act.

9.1.3 Data on vaccination carried out

Vaccination against avian influenza is prohibited and vaccination has not been performed before.
10. Description of the epidemiological situation of the disease in wild birds during the last five years

(max. 4000 chars):

Within the surveillance of wild birds there were tested in total 653 birds in 2010. In the frame of active surveillance, there were tested 500 hunted wild birds (mallards) whereas 76 mallards of them were virological positive for LPAI (subtype H5 and H7 were negative). Other 153 wild birds were examined for avian influenza in the frame of passive surveillance (found dead). Only one bird (mallard) was positive for LPAI subtype H1N9. In total there were 77 positive wild birds for LPAI.

Within the surveillance of wild birds there were tested 751 birds in 2009. Out of this number were tested 532 birds (mallards – 4 clinically health and 528 hunted) in the frame of active surveillance and 219 birds (found death) within passive surveillance. In total 25 birds were positive.

There were 814 wild birds tested in 2008. In the frame of passive surveillance there were examined 259 wild birds (found dead) and in the frame of active surveillance were tested 555 wild birds (category hunted without clinical signs: 551 birds, category live without clinical signs: 4 birds). In total there were 57 positive wild birds for LPAI from what five birds were positive for LPAI H5N3. H5 and H7 subtypes were ruled out in other 52 cases.

Within the surveillance in wild birds, 649 wild birds in total were examined during the period from 1 January 2007 to 31 December 2007 and one positive virological finding was detected in wild swan in South-Moravian Region (catastral territory of municipality Lednice) on 25th June 2007.

The first occurrence of the High Pathogenic Avian Influenza (HPAI) on the Czech Republic territory was detected in the year 2006. HPAI was detected only in wild birds. In total 14 cases were found in the species “Mute swan”. In all these 14 cases the high pathogenic subtype H5N1 was confirmed. All cases were reported in birds which were found dead in the South Bohemian and South Moravian regions.

10.1 Measures included in the programme for surveillance in wild birds
Measures included in the programme for surveillance in wild birds are described in three following articles 10.1.1., 10.1.2. and 10.1.3.

10.1.1 Designation of the central authority in charge of supervising and coordinating the departments responsible for implementing the programme

The central authority competent for supervising and coordinating all activities in veterinary care is State Veterinary Administration, which performs its powers at the whole territory of the Czech Republic (§ 47, Veterinary Act No 166/1999 Col. of Acts). SVA of the CR coordinates the activities of Regional Veterinary Administrations (RVA). The surveillance programme for AI is laid down on the base of § 48 point 1, and § 10, Veterinary Act No. 166/1999 as amended.

Ministry of Agriculture of the Czech Republic determines main strategies in a veterinary care and carries out their control as laid down in the Veterinary Act No. 166/1999 Article 44, Point 1 a. The Ministry of Agriculture specifies obligatory preventive and diagnostics campaigns in accordance with the Veterinary Act, Article 44; Point 1d, based on the epidemiological situation. Related details are laid down in the “Methodology of Animal Health Controls and Prophylaxis” approved by the Ministry of Agriculture and issued in its Official Journal. According to the legislation (Veterinary Act 166/1999) the SVA CR has the legal power to supervise any action ordered by the “Methodology”. Regional veterinary administrations execute the legal powers as to supervise private veterinarians over their actions in the professional field as ordered by the “Methodology”.

The competent authority confirms its commitment to submit a complete report on actions and expenditures in the framework of this programme and to provide additional information when they are request by the Commission.
10.1.2  Description and delimitation of the geographical and administrative areas in which the programme is to be applied

(max. 4000 chars):

The programme will be applied in all territory of the Czech Republic according to rules setting by the state veterinary administration.

10.1.3  Estimation of the local and/or migratory wildlife population

(max. 4000 chars):

The Czech Republic is out of the main migratory flight paths of birds from central and eastern Asia, the Caspian Sea and the Black sea areas. Due to reason according the epidemiological and ornithological data we have not identified any region at higher risk for the introduction of AI. There are identified only regions with the large waters areas where the majority of waterfowl nest.

11. Measures in place as regards the notification of the disease

(max. 4000 chars):

12. Costs

12.1 Detailed analysis of the costs

12.1.1 Poultry

The overall estimated cost of the AI surveillance in poultry during year 2012 is 25 205.2 €. The estimated number of 225 poultry holdings is to be tested in the frame of AI surveillance programme during 2012. The cost of the programme including the cost of a lump sum of 0,5 € per poultry sampled, laboratory testing by pre-screening serological tests and cost for following tests in case of positive pre-screening test. The competent authority wishes 50% of co-financing of the cost to be incurred for the costs of carrying out laboratory tests and a lump sum of 0,5 € per poultry sampled as the financial contribution of the Union.

12.1.2 Wild birds

The overall estimated cost of the AI surveillance in wild birds during year 2012 is 41 720 €. The estimated number of 300 wild birds is to be tested in the frame of AI surveillance programme during 2012 (passive surveillance on sick and dead wild birds). The cost of the programme including the cost of a lump sum of 5 € per wild bird sampled and virological laboratory testing. The competent authority wishes 50% of co-financing of the cost to be incurred for the costs of carrying out laboratory tests and a lump sum of 5 € per wild bird sampled as the financial contribution of the Union.
### 12.2 Summary of the costs

#### 12.2.1 Poultry surveillance

<table>
<thead>
<tr>
<th>Methods of laboratory analysis</th>
<th>Number of tests to perform per method</th>
<th>Unitary test cost (per method) in €</th>
<th>Total cost (€)</th>
</tr>
</thead>
<tbody>
<tr>
<td>ELISA test</td>
<td>3 070</td>
<td>4.48</td>
<td>13 753.6</td>
</tr>
<tr>
<td>agar gel immune diffusion test</td>
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<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Haemagglutination-inhibition-test (HI) for H5</td>
<td>300</td>
<td>4.48</td>
<td>1 344</td>
</tr>
<tr>
<td>Haemagglutination-inhibition-test (HI) for H7</td>
<td>300</td>
<td>4.48</td>
<td>1 344</td>
</tr>
<tr>
<td>Virus isolation test</td>
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<td>35.42</td>
<td>709</td>
</tr>
<tr>
<td>PCR test</td>
<td>80</td>
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<td><strong>3 770</strong></td>
<td></td>
<td><strong>20 410.60 €</strong></td>
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</tbody>
</table>

**Other measures to be covered**

<table>
<thead>
<tr>
<th>Measures to be covered</th>
<th>Number of tests to perform</th>
<th>Unitary test cost (per method) in €</th>
<th>Total cost (€)</th>
</tr>
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<tr>
<td>Sequencing H cleavage site</td>
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<tr>
<td>Sequencing N cleavage site</td>
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<td>1 629.8</td>
</tr>
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<td>1 535</td>
</tr>
</tbody>
</table>
### Standard requirements for the submission of surveillance programmes for avian influenza

**version : 2.1**

<p>| | | | | | |</p>
<table>
<thead>
<tr>
<th></th>
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<td></td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td>163</td>
<td></td>
<td></td>
<td></td>
<td>4 794,60 €</td>
</tr>
</tbody>
</table>

Add a new row
### 12.2.2 Wild bird surveillance

<table>
<thead>
<tr>
<th>Methods of laboratory analysis</th>
<th>Number of tests to perform per method</th>
<th>Unitary test cost (per method) in €</th>
<th>Total cost (€)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Haemagglutination-inhibition-test (HI) for H5/H7</td>
<td>0</td>
<td>0</td>
<td>0</td>
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<tr>
<td>Virus isolation test</td>
<td>100</td>
<td>35.45</td>
<td>3545</td>
</tr>
<tr>
<td>PCR test</td>
<td>300</td>
<td>40.75</td>
<td>12225</td>
</tr>
<tr>
<td>Sequencing H and N cleavage site</td>
<td>150</td>
<td>163</td>
<td>24450</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td><strong>400</strong></td>
<td><strong>76,20 €</strong></td>
<td><strong>40 220,00 €</strong></td>
</tr>
</tbody>
</table>

#### Other measures to be covered

<table>
<thead>
<tr>
<th>Sampling</th>
<th>Number of tests to perform per method</th>
<th>Unitary test cost (per method) in €</th>
<th>Total cost (€)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>300</td>
<td>5</td>
<td>1500</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td></td>
<td></td>
<td><strong>1 500,00 €</strong></td>
</tr>
</tbody>
</table>

Add a new row
Attachments

IMPORTANT:
1) The more files you attach, the longer it takes to upload them.
2) This attachment files should have one of the format listed here: .zip, .jpg, .jpeg, .tiff, .tif, .xls, .doc, .bmp, .pna.
3) The total file size of the attached files should not exceed 2 500Kb (+- 2.5 Mb). You will receive a message while attaching when you try to load too much.
4) IT CAN TAKE SEVERAL MINUTES TO UPLOAD ALL THE ATTACHED FILES. Don't interrupt the uploading by closing the pdf and wait until you have received a Submission Number!
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