

**APPENDIX 4: EUROPEAN BASELINE STUDY FOR
CRY1AB SUSCEPTIBILITY IN
OSTRINIA NUBILALIS IN THE 2009
SEASON**

([REDACTED] 2010. Baseline study of
Cry1Ab susceptibility in populations of
Ostrinia nubilalis (ECB) – Results for
2005-2009. Sagerheide, Germany.)

Report

Baseline study of Cry1Ab susceptibility in populations of *Ostrinia nubilalis* (ECB)

- Results for 2005-2009 -

Date

13/08/2010

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**Statement of Compliance with the Principles of
Good Experimental Practice**

The study described in this report was conducted in compliance with the most recent edition of:

- The Principles of Good Experimental Practice (GEP), (Plant Protection Products Ordinance, paragraph (5) of Article 1c, Germany).

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1. Introduction

The European corn borer (ECB), *Ostrinia nubilalis*, is native to southern Europe (BECK, 1987) and is believed to have been introduced into North America between 1909 and 1914 (VINAL, 1917), where multiple introductions have probably occurred (SHOWERS, 1993). Since then ECB has rapidly spread across North America (CAFFREY and WORTHLEY, 1927; ROELOFS et al., 1985; HUDON & LEROUX, 1986). Apart from maize, more than 200 weeds and cultivated plants are known to serve as host plants for ECB (HODGSON, 1928; PONSARD et al., 2004). Now ECB is one of the most damaging pests of maize in North America and Europe and a major target pest for control with transgenic Bt corn.

In order to maintain the benefits obtained from growing *B. thuringiensis* (Bt) maize varieties, Monsanto established an insect monitoring program across Europe and in particular in areas where commercial activity of MON 810 genetically modified maize is occurring or planned. An important need prior to the growing of Bt maize varieties consists of establishing the baseline susceptibility of field populations of ECB to the Bt Cry1Ab protein, which is the active ingredient in MON 810. Then, every two years, a routine monitoring program will survey and quantify any potential change in susceptibility in ECB field populations exposed to Bt maize cultivation. This program will enable early detection of resistance in ECB if it occurs, and this will allow the proposal and implementation of additional risk mitigation measures. For efficiency, the baseline studies and monitoring of resistance have been focused in areas where the potential resistance risk is relatively high because the introduction of Bt maize is relatively high (or expected to be), and local entomologists recognize ECB to be abundant.

Previous baseline susceptibility studies. Baseline susceptibility to Cry1Ab protein has been established for ECB populations collected in different maize areas in Spain (GONZALEZ-NUNEZ et al., 2000, FARINÓS et al., 2004), Germany (SAEGLITZ et al., 2006) and the United States (MARÇON et al., 1999a and 2000). The methodology for those studies involved applying a solution with the appropriate Cry1Ab protein concentrations on the surface of artificial diet (GONZALEZ-NUNEZ et al., 2000).

2. Materials and Methods

2.1. Insect Collection

For the current study, ECB were collected from 2005–2009 in major European maize growing regions: Czech Republic/Moravia (CZ), Southwest and West France (Fsw, Fw), Northern Germany/Southwest Poland (GnPLw), Southern Germany and East France (GsFe), Northern Italy (ITne, ITnw), the Panonian region (PAN, Western Slovakia and North West Hungary), Southeast Poland (PLse), South Portugal (Ps), Romania (ROe, ROW) and Spain (ESne, ESc, ESsw; see Fig. 1). For each region, different sampling sites separated by at least 50 to 100 km were chosen. ECB were collected as adults, larvae or egg masses in naturally infested fields. Larvae were collected by dissecting corn stalks a few days before harvesting or spring after diapause. If more than one larva per stalk was found, only one was taken to avoid collecting siblings (Fig. 2). For each sampling site, the aim was to collect at least 300 larvae (Table 1).

To collect egg masses, light traps and large cages (2 x 2 x 2 m) were used in corn fields. Adult ECB were attracted by the light during flying season and caught alive in the cages. Every two to three days, egg masses oviposited on the corn plants in the cage were cut off and transferred to the laboratory. The number of collected egg masses varied between 35 and 815.

This insect collection and population establishment scheme is in compliance with the industry IRM (Insect Resistance Management) working group guidelines proposed to the competent authority (EU Commission).

2.2. Insect Culture

Field-collected ECB larvae were placed in plastic boxes containing corrugated cardboard and maintained in a growth chamber at 25°C, 90% RH and a photoperiod of 20:4h (L:D) on an agar-based wheat germ diet (Table 2). If the larvae did not pupate after a period of two weeks, they were assumed to have entered diapause and were transferred to another climatic chamber maintained at $8 \pm 2^\circ\text{C}$, $70 \pm 5\%$ RH, and a photoperiod of 0:24h (L:D) (Fig. 3). All ECB collected as larvae (L3–L5) entered diapause after collection. The mass of larvae varied between 61–165 mg at the beginning of diapause.

In early 2006 it was tested if diapause can be broken artificially. After three months in diapause conditions a portion of the larvae collected near Heinersdorf/Oderbruch, Germany (G.03) were placed at $25 \pm 2^\circ\text{C}$, $80 \pm 5\%$ RH, and a photoperiod of 24:0h (L:D). The cardboard was moistened daily until pupation. The emergence of these adults occurred over a period of nearly two months at a very low level. This complicated the production of a sufficient number of egg masses to be used in subsequent bio assays. To achieve a more synchronized adult eclosion it was therefore decided to allow the completion of diapause until May (as in nature) as standard practice.

Larvae surviving the diapause period were transferred to fresh containers to prevent contamination by dead larvae. To increase the temperature step-wise, these containers were maintained for 5d at $15 \pm 2^\circ\text{C}$, for the next 5d at $20 \pm 2^\circ\text{C}$, and thereafter at $25 \pm 2^\circ\text{C}$. At all temperature steps, larvae were maintained at $90 \pm 5\%$ RH and a photoperiod of 24:0h (L:D). During this time, additional water for drinking was added. Any food supplied was not taken by the larvae. To transfer emerged adults from diapause boxes into oviposition cages CO_2 was used.

Egg masses were obtained by confining batches of up to ten pairs of adults in cylindrical plastic tubes (\varnothing 11 cm, Fig. 5). Adults were fed with 15 % honey-water to increase fecundity and egg laying (LEAHY & ANDOW, 1994). The inside of the tubes were covered with filter paper as oviposition medium. These paper sheets were changed twice a week. Egg masses were cut off and transferred to petri dishes with moistened filter paper. If necessary, egg masses were stored for up to seven days at $8 \pm 2^\circ\text{C}$. Oviposition cages and incubated egg masses were maintained in an environmental chamber for 20 h at $25 \pm 2^\circ\text{C}$, 4 h at $20 \pm 2^\circ\text{C}$, $90 \pm 5\%$ RH and a photoperiod of 20:4h (L:D) (GUTHRIE et al., 1985).

2.3. Bioassays

Cry1Ab toxin was provided by Monsanto and was stored at -82°C until used. To prepare the test concentrations, a bicarbonate buffer (0.25 mmol/l) with pH 10.5 was used. The bioassays were performed in 128 well trays (Bio-Ba-128, Color-Dec, Italy). In each cell 1 ml of artificial diet was dispensed. After the diet solidified, 100 μl of toxin solution was applied to the surface and allowed to dry over night at room temperature. To avoid contaminations the trays were covered with a sheet of filter

paper. Egg masses of each sampling location (field-collected or offspring of collected larvae) were incubated and neonate larvae, within 12 h after hatching, were transferred to the cells. If the number of collected egg masses was too low to perform an assay, ECB were reared for one generation and the resulting offspring were tested. A single neonate was placed in each cell and confined with a cover (Bio-Cv-16, Color-Dec Italy) (Fig. 6). Eight concentrations (0.5–256.0 ng Cry1Ab/cm²) and a control (bicarbonate buffer) were tested for each population. Each concentration was tested with at least 16 larvae. All assays were conducted at 25°C, 70% RH and a photoperiod of 0:24h (L:D). After seven days, larval mortality and developmental stage were recorded. Larvae that had not grown beyond first instar were considered to be dead because larvae unable to moult under field conditions would not survive (e.g. SIEGFRIED et al., 2000). As a result, the criterion for mortality used in this study accounts for both death and complete moulting (or growth) inhibition.

2.4. Statistical Analysis

All statistical analyses were done using the computer programme SYSTAT, Version 10.0, except for concentration-response analysis where PoloPlus 1.0 was used (LeOra Software Company). The measure of how well the data (response of ECB to different concentrations of toxin) fit the assumptions of the probit model is goodness-of-fit. To test goodness-of-fit, responses predicted by the probit model were compared with responses actually observed in the bioassay (χ^2 test).

Hypothesis tests are essential for the interpretation of bioassay results. Three possible outcomes of comparing probit regression lines are that lines are parallel but not equal, lines are equal, or lines are neither parallel nor equal. When lines are parallel but not equal, their slopes are not significantly different. This means that changes in activity per unit change in rate are the same. If regression lines are equal, they do not differ in either intercept or slope, meaning the populations being compared are equally affected. When lines are neither equal nor parallel, neither their intercepts nor their slopes are equal, meaning the populations being compared differ in susceptibility and response characteristics.

Ratios of activity at a response level such as 50 or 90% provide a means to estimate the relative susceptibility of populations of ECB to Cry1Ab. To provide an estimate of the variability involved in these ratios, 95% confidence intervals are calculated for

each ratio. To determine whether the response of one group differs significantly from the other, their 95% confidence limits are compared. If the limits overlap, then the lethal concentrations do not differ significantly (at an error rate of 5%).

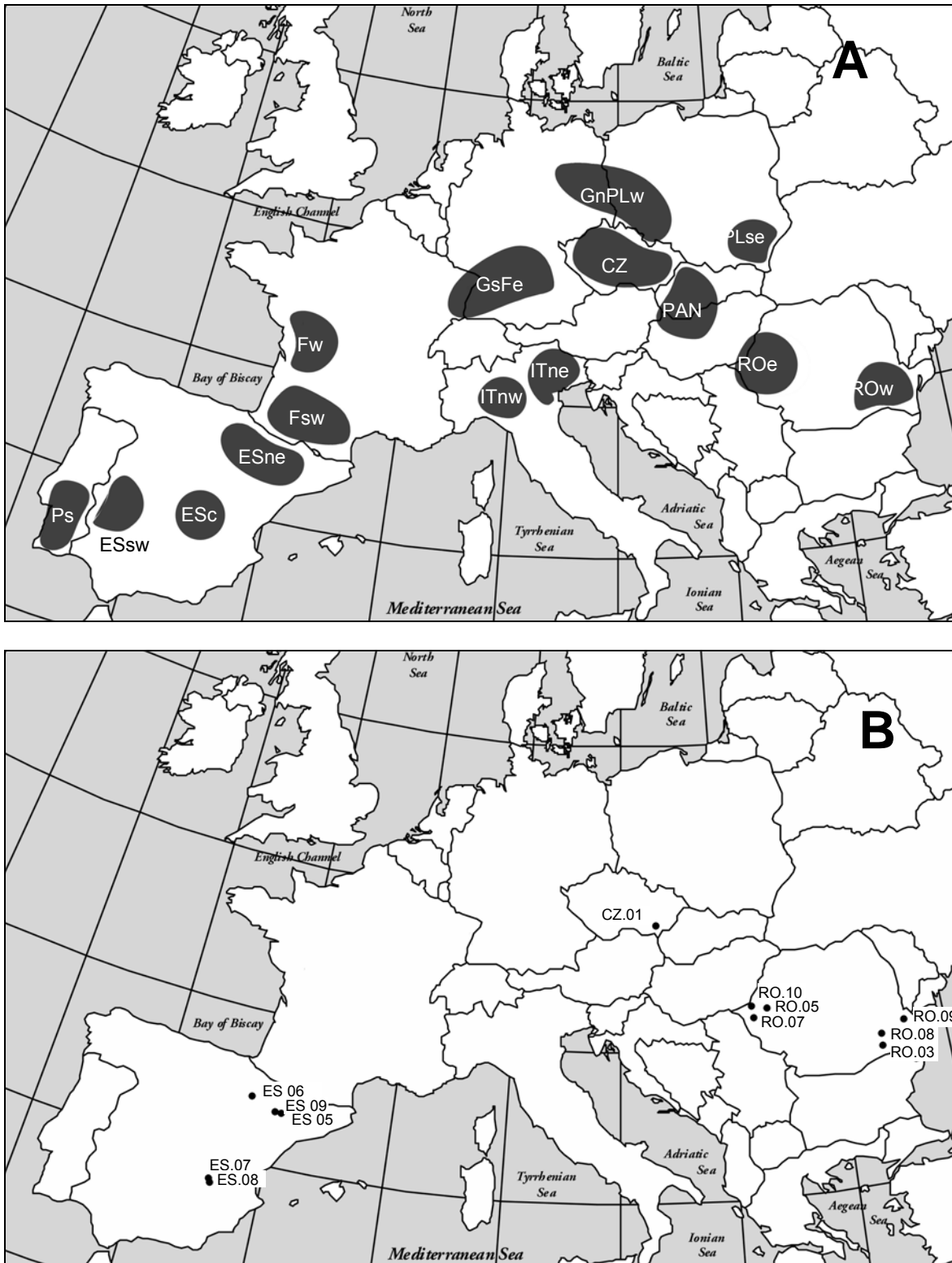


Fig. 1: ECB populations considered in Europe (A) and sites sampled 2009 (B)



Fig. 2: Dissected maize stalk with larvae



Fig. 3: Growth chamber with plastic boxes containing diapausing ECB larvae



Fig. 4: Corrugated cardboard with pupae



Fig. 5: Oviposition cages for adult ECB

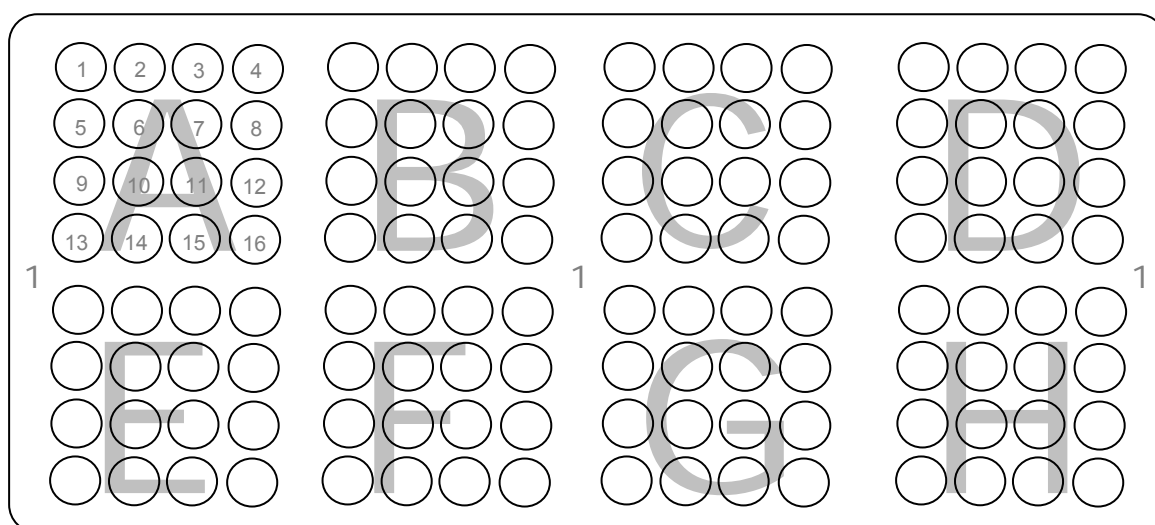


Fig. 6: IDs of Bio-Ba-128 trays (tray number, field letter, well number; i.e.: 1.A.13)

Table 1: Source description of ECB collections used to establish baseline susceptibility to the Cry1Ab toxin (in prep. – in preparation, results will be reported 2011).

Country	Collection site	ID	Population	Collected	Eggs	Larvae	Adults	Tested	
Czech	Čejč	CZ 01	CZ	2005		x		2006	
	Velké Bílovice	CZ 01	CZ	2009		x		2009	
	Ivanovice na Hané	CZ 02	CZ	2005		x		2006	
		CZ 02	CZ	2008	x			2008	
	Klapy	CZ 03	CZ	2005			x		2006
		CZ 03	CZ	2007			x		dead
CZ 03		CZ	2008	x				2008	
France	Wiwersheim	F 01	GsFe	2005	x			dead	
		F 01	GsFe	2006		x		2007	
		F 01	GsFe	2007	x			2007	
	Miradoux dans le Gers	F 02	Fsw	2005			x		2006
		F 02	Fsw	2008			x		2009
	Berat en Haute-Garonne	F 03	Fsw	2005			x	2006	
	Lezat Sur Leze en Ariège	F 04	Fsw	2005			x	2006	
	Savigny	F 05	Fw	2006	x			2006	
	Pamproux	F 06	Fw	2006	x			2006	
	Taizé-Aizié	F 07	Fw	2006	x			2006	
	Saint Barthelemy	F 08	Fsw	2006			x		2007
		F 08	Fsw	2007			x		2008
		F 08	Fsw	2008			x		dead
	Auriac	F 09	Fsw	2006			x	2007	
	Solomiac (Tarn et	F 10	Fsw	2006			x	2007	
	Frégouville	F 11	Fsw	2007			x	dead	
Lavaur	F 12	Fsw	2007			x	dead		
Orgueil	F 13	Fsw	2008			x	2009		
Saint Bonnet sur-Gironde	F 14	Fw	2008			x	dead		
Lusignan	F 15	Fw	2008			x	dead		
Thurageau	F 16	Fw	2008			x	dead		
Germany	Herbolzheim	G 01	GsFe	2005	x	x		2006	
		G 01	GsFe	2007			x	dead	
	Sugenheim	G 02	GsFe	2005			x	2006	
	Heinersdorf	G 03	GnPL	2005			x	2006	
		G 03	GnPL	2008	x			2008	
	Niedernberg (reference-strain)	G 04	lab	2005	x			2006	
		G 04	lab	2005	x			2007	
		G 04	lab	2005	x			2008	
	Ansbach	G 05	GsFe	2006	x			2006	
	Keindorf	G 06	GnPL	2006			x	2007	
G 06		GnPL	2008			x	dead		
Altreetz	G 07	GnPL	2007	x			2007		
	G 07	GnPL	2008	x			2008		
Ulsenheim/Uffenheim	G 09	GsFe	2007	x			2007		
Kitzingen	G 10	GsFe	2007			x	2008		
Hungary	Bana/Gulyaré	HU 01	PAN	2006			x	2006	
		HU 02	PAN	2006			x	dead	

Table 1: continued

Country	Collection site	Site ID	Population	Collected	Eggs	Larvae	Adults	Tested
Italy	Cicignolo	IT 01	ITnw	2007		x		dead
	Ghisalba	IT 02	ITnw	2007		x		dead
	Belfiore	IT 03	ITnw	2007		x		dead
	Fiume Veneto	IT 04	ITne	2007		x		dead
	Gonars	IT 05	ITne	2007		x		dead
	Gonars	IT 06	ITne	2007		x		dead
	Padova	IT 07	ITne	2008		x		2008
	Vicenza	IT 08	ITne	2008		x		2008
	Fossalta di Piave	IT 09	ITne	2008		x		2008
Poland	Rzeszow	PL 01	PLse	2005			x	dead
	Rynakowice	PL 02	GnPLw	2006		x		dead
		PL 02	GnPLw	2007		x		2008
		PL 02	GnPLw	2008	x			2008
	Krzeczowice	PL 03	PLse	2006		x		dead
		PL 03	PLse	2007		x		2008
		PL 03	PLse	2008	x			2008
Portugal	Mix of three sites	P 01	Ps	2008	x		2009	
Slovakia	Levice	SK 01	PAN	2006	x			2006
		SK 01	PAN	2008	x			2008
	Vrbové	SK 02	PAN	2006	x			2006
	Trnava	SK 03	PAN	2008	x			2008
Spain	Hernán Cortés	ES 01	ESsw	2008		x		2009
	Don Benito	ES 02	ESsw	2008		x		2009
	Obando	ES 03	ESsw	2008		x		2009
	Gimenells	ES 04	ESne	2008		x		2009
	Candasnos	ES 05	ESne	2008		x		dead
		ES 05	ESne	2009		x		2010
	Erla/ Santa Anastasia	ES 06	ESne	2008		x		2009
	Ejea de los Caballeros	ES 06	ESne	2009		x		2010
	Barrax/Aguas Nuevas	ES 07	ESc	2008		x		dead
	Aguas Nuevas	ES 07	ESc	2009		x		2010
	El Salobral	ES 08	ESc	2009		x		2010
Bujaraloz	ES 09	ESc	2009		x		2010	
Romania	Simand	RO 01	ROw	2007		x		dead
	Fundulea	RO 02	ROe	2007		x		dead
		RO 02	ROe	2008		x		2009
		RO 03	ROe	2008		x		dead
	Cascioarele	RO 03	ROe	2009				2010
	Dalga	RO 04	ROe	2008		x		dead
	Paulis	RO 05	ROw	2008		x		2009
	Lipava	RO 05	ROw	2009		x		in prep.
	Buteni	RO 06	ROw	2008		x		2009
	Sandra	RO 07	ROw	2008		x		2009
	Carpinis	RO 07	ROw	2009		x		in prep.
	Berceni	RO 08	ROe	2009		x		2010
	Tudor Vladimirescu	RO 09	ROe	2009		x		2010
Nadlac	RO 10	ROw	2009		x		2010	

Table 2: Artificial diet for rearing ECB

Water	680 ml
Benzoic acid ²	1 g
Sorbic acid ¹	1 g
Nipagin (methyl-paraben) ¹	1 g
Agar-agar ^{*2}	16 g
Maize powder ³	112 g
Wheatgerm ⁴	28 g
Brewer's yeast ⁵	30 g
Fumidil B ⁶	1 g
Ascorbic acid ¹	3 g
Vanderzant vitmain mix ¹	2 g

* To prepare diet for Cry1Ab bioassays the amount of agar was halved to 8 g.

1. BioServ; One 8th Street, NJ 08825 Frenchtown, USA

Prod.No. 6030 (ascorbic acid)

Prod.No. 6967 (sorbic acid)

Prod.No. F8045 (vitamin mix)

Prod.No. 7685 (nipagin, methyl-paraben)

2. Carl Roth GmbH & Co. KG; Schoemperlenstr. 3–5, 76185 Karlsruhe, Germany

Art.No. 5210.2 (Agar-Agar, Kobe 1 pulv.)

Art.No. 5781.1 (Benzoic acid, ≥ 99.5 %)

3. Gut & Gerne, BZ Bio-Zentrale GmbH; 94166 Stubenberg, Germany

4. Frießinger Mühle GmbH; 74206 Bad Wimpfen, Germany

5. Biolabor GmbH & Co.KG; PF 15 01 31, 28091 Bremen, Germany

6. CEVA Salud Animal, S.A., c. Carabela La Nina, 12 5^a planta, 08017 Barcelona, Spain

3. Results and Discussion

Bioassays estimating parameters such as the LC_{50} or LC_{90} are recommended as part of resistance management strategies (SIMS et al., 1996). It is most convenient to conduct the bioassays with purified toxin comparable to that produced by the Bt plant.

Using artificial diet, the toxin can be provided as a surface treatment (GOULD et al., 1997; HILBECK et al., 1998) or mixture (SAEGLITZ et al., 2006).

Susceptibility data for ECB collected in different geographic regions and exposed to purified Cry1Ab toxin are presented in Table 3, Figure 7 and appendix Figures A01-14. The lowest MIC_{50} value (1.20 ng/cm^2) was found for a colony collected 2002 in Niedernberg, Germany (G 04) and kept since then as laboratory strain. Repeated analysis showed small MIC variability for this strain within a season and between years. The differences between the smallest and greatest MIC_{50} were 3.0-fold and for MIC_{90} 2.4-fold (Tab. 3). For the field-collected ECB, the MIC_{50} values ranged from 5.71 ng/cm^2 (Ivanovice na Hané, CZ 02) to 22.37 ng/cm^2 (Sugenheim, G 02) in 2005, 2.45 ng/cm^2 (Wiwersheim, F 01) to 20.33 ng/cm^2 (Savigny, F 05) in 2006, 3.44 ng/cm^2 (Rynakowice, PL 02) to 9.08 ng/cm^2 (Saint Barthelemy F 08) in 2007, 1.71 ng/cm^2 (Ivanovice na Hané, CZ) to 14.29 ng/cm^2 (Fundulea RO 02) in 2008, and from 2.39 ng/cm^2 (El Salobral ES 08) to a maximum of 13.56 ng/cm^2 (Cascioarele, RO 03) in 2009. Differences between the most susceptible and most tolerant field-collected samples were 13.1-fold for all years. In 2009 the MIC_{50} variation was 5.7-fold.

Results for populations pooled according to geographic and climatic conditions are presented in Table 4. Populations pooled correspond to homogeneous regions based on available knowledge of insect biology and geography. This approach follows the IRM industry working group guidelines.

Although variation in susceptibility to Cry1Ab was found among samples and among regions, the magnitude of the variation in MIC_{50} was small (i.e. 3.9-fold, 8.3-fold, 2.6-fold, 8.4-fold, and 5.7-fold for ECB collected in 2005, 2006, 2007, 2008, and 2009 respectively, Tabl. 3). The results of the populations pooled according to geographic and climatic conditions were similar; the MIC_{50} values differed 1.8-fold, 6.6-fold, 2.6-fold, 4.2-fold and 3.2-fold for ECB collected in 2005, 2006, 2007, 2008, and 2009 respectively (Tabl. 4). A similar degree of variability was reported for ECB susceptibility to Cry1Ab for populations from three broad geographic areas in the US, chosen based on market penetration for Bt corn (Siegfried et al., 2007). Similar levels

of variability were also observed in a study that included populations of different voltine ecotypes and pheromone strains (Marçon et al., 1999b). For the current study, the pheromone races were not distinguished.

Table 3. Susceptibility of ECB neonates exposed to Cry1Ab. (¹ year tested, collected 2002, * - no concentration response relationship; MIC - moultin inhibition concentrations estimated by probit analysis PoloPlus (LeOra Software, 1987); CI - confidence interval; ^a - ng/cm²)

Population	Collected	Site ID	MIC ₅₀ (95% CI) ^a	MIC ₉₀ (95% CI) ^a
Reference	2006 ¹	G 04	1.20 (0.50–2.21)	4.78 (2.57–14.38)
	2007 ¹	G 04	1.44 (0.86–2.06)	3.94 (2.68–8.28)
	2008 ¹	G 04	2.21 (1.89–2.55)	4.47 (3.70–6.00)
	2008 ¹	G 04	2.26 (1.49–3.01)	8.16 (5.95–13.50)
	2009 ¹	G 04	3.65 (2.77–4.90)	9.56 (6.72–17.75)
Czech Republic	2005	CZ 01	8.22 (6.31–10.75)	19.53 (14.25–33.61)
		CZ 02	5.71 (4.03–7.76)	18.40 (12.61–35.73)
		CZ 03	7.42 (6.09–8.67)	12.26 (10.28–16.87)
	2008	CZ 02	1.71 (0.43–3.04)	6.97 (3.86–39.16)
		CZ 03	9.04 (6.40–13.35)	20.79 (13.94–46.82)
	2009	CZ 01	9.00 (5.74–12.35)	17.67 (12.80–37.88)
France Southwest	2005	F 02	*	
		F 04	*	
		F 03	7.26 (1.87–12.69)	28.35 (16.66–75.72)
	2006	F 08	3.21 (2.42–3.85)	5.66 (4.67–7.97)
		F 09	2.65 (1.90–3.35)	5.05 (3.88–9.80)
		F 10	2.49 (1.66–3.20)	5.07 (3.83–10.49)
	2007	F 08	9.08 (4.75–12.92)	26.74 (18.13–68.38)
	2008	F 02	4.43 (2.80–6.90)	10.69 (6.87–30.28)
		F 13	5.38 (3.89–6.99)	9.59 (7.30–20.84)
	France West	2006	F 05	20.33 (17.74–23.31)
F 06			*	
F 07			3.86 (1.56–6.20)	14.35 (8.94–35.27)
Germany North/ Poland West	2005	G 03	13.33 (9.45–17.64)	35.70 (25.97–60.49)
	2006	G 06	2.82 (1.87–3.70)	6.60 (4.84–13.18)
		G 07	6.88 (5.80–7.84)	10.14 (8.75–13.76)
	2007	PL 02	3.44 (2.69–4.38)	6.93 (5.27–11.50)
		G 03	3.13 (2.47–3.95)	5.99 (4.62–9.78)
		PL 02	3.77 (2.89–4.92)	8.11 (5.98–14.38)
2008	G 07	4.45 (2.90–5.96)	8.73 (6.43–19.19)	

Table 3 continued

Population	Collected	Site ID	MIC ₅₀ (95% CI) ^a	MIC ₉₀ (95% CI) ^a
Germany South/ France East	2005	G 01	6.26 (3.52–9.80)	24.95 (14.90–71.84)
		G 02	22.37 (17.38–27.02)	30.32 (25.29–43.10)
	2006	F 01	2.45 (1.83–3.05)	3.72 (3.00–6.52)
		G 05	7.81 (2.04–12.96)	27.54 (16.91–81.40)
	2007	F 01	4.63 (3.16–6.21)	10.94 (7.88–21.09)
		G 09	5.50 (4.10–7.29)	12.95 (9.38–23.30)
G 10		8.07 (5.43–11.22)	23.54 (16.08–47.96)	
Italy Northeast	2008	IT 07	8.43 (6.96–9.71)	18.09 (14.76–24.38)
		IT 08	11.97 (9.42–15.30)	20.61 (15.99–33.59)
		IT 09	11.04 (8.28–14.72)	27.55 (19.77–47.80)
Panonia	2006	HU 01	8.29 (6.60–10.25)	13.32 (10.68–21.29)
		SK 02	3.94 (1.16–6.99)	19.97 (11.54–55.74)
		SK 01	4.51 (2.00–7.24)	18.78 (11.58–45.49)
	2008	SK 01	4.19 (2.97–5.49)	8.51 (6.31–17.51)
		SK 03	3.98 (2.99–5.30)	10.45 (7.43–18.63)
	2010	SK 01	8.24 (2.80–33.08)	40.74 (14.52–2175.76)
		SK 0y	8.92 (5.44–12.28)	20.03 (14.23–45.12)
Poland Southeast	2007	PL 03	6.29 (3.49–12.48)	15.86 (8.96–86.93)
	2008	PL 03	*	
Portugal South		P 01	3.66 (2.46–4.78)	11.90 (8.80–20.04)
Romania East	2008	RO 02	14.29 (10.56–19.19)	38.18 (27.16–65.65)
	2009	RO 03	13.56 (9.72–18.38)	34.96 (24.74–63.37)
		RO 08	6.38 (3.18–12.43)	13.66 (8.18–96.60)
		RO 09	11.93 (5.70–17.73)	35.67 (23.63–87.00)
Romania West	2008	RO 05	9.73 (6.81–13.26)	25.06 (17.67–46.67)
		RO 06	6.63 (3.51–10.73)	16.16 (10.13–65.51)
		RO 07	9.54 (7.09–12.87)	23.64 (16.79–42.63)
	2009	RO 10	7.73 (3.50–22.49)	21.11 (10.36–422.61)
Spain Central	2009	ES 07	4.22 (2.94–5.51)	9.71 (7.17–17.90)
		ES 08	2.39 (1.19–4.08)	11.85 (6.55–36.55)
Spain Northeast	2008	ES 04	7.21 (3.52–13.77)	21.34 (11.65–102.29)
		ES 06	6.83 (3.79–12.20)	26.09 (14.15–95.20)
	2009	ES 05	7.91 (6.00–10.69)	19.87 (14.01–36.25)
		ES 06	7.45 (5.87–9.84)	14.58 (10.78–27.13)
		ES 09	4.25 (3.29–5.34)	7.33 (5.74–12.70)
Spain Southwest	2008	ES 01	2.83 (2.19–3.68)	6.31 (4.66–10.84)
		ES 02	3.67 (2.30–5.41)	6.53 (4.63–19.11)
		ES 03	3.71 (2.89–4.77)	7.63 (5.74–13.14)

Table 4. Susceptibility of ECB neonates exposed to Cry1Ab as measured by the MIC, a combination of growth inhibition and mortality. Collections were pooled according to geographic and climatic similarity. (^a ng Cry1Ab/cm²; MIC moulting inhibition concentrations, CI confidence interval, * no concentration response)

Population	MIC50 (95% CI) ^a	MIC90 (95% CI) ^a
Czech Republic		
2005 (CZ 01, CZ 02, CZ 03)	7.59 (6.30–8.72)	15.47 (13.16–19.89)
2008 (CZ 02, CZ 03) ¹	*	
2009 (CZ 01)	8.99 (5.74–12.35)	17.67 (12.80–37.88)
France Southwest		
2005 (F 02, F 03, F 04)	11.13 (1.38–25.78)	56.98 (24.50–296.54)
2006 (F 08, F 09, F 10)	3.14 (2.58–3.61)	5.49 (4.74–6.91)
2007 (F 08)	9.08 (4.75–12.92)	26.74 (18.13–68.38)
2008 (F 02, F 13)	4.81 (3.89–5.88)	10.70 (8.37–16.01)
France West		
2006 (F 05, F 06, F 07)	18.48 (15.53–21.67)	25.97 (22.06–37.29)
Germany North/Poland West		
2005 (G 03)	13.33 (9.45–17.64)	35.70 (25.97–60.49)
2006 (G 06)	2.82 (1.87–3.70)	6.60 (4.84–13.18)
2007 (G 07, PL 02)	5.58 (4.04–7.53)	11.87 (8.58–22.98)
2008 (G 03, G 07, PL 02)	3.80 (1.82–7.28)	9.26 (5.34–61.55)
Germany South/France East		
2005 (G 01, G.02) ¹	**	
2006 (F 01, G 05)	4.79 (2.42–7.43)	20.00 (12.44–48.41)
2007 (F 01, G 09, G 10)	5.78 (4.84–6.79)	14.88 (12.11–19.80)
Italy Northeast		
2008 (IT 07, IT 08, IT 09)	9.47 (8.26–10.80)	21.10 (17.81–26.44)
Panonia		
2006 (HU 01, SR 01, SR 02)	6.13 (2.83–8.76)	18.33 (13.33–32.59)
2008 (SK 01, SK 03)	4.10 (3.24–4.89)	9.87 (7.67–14.62)
2010 (SK 01 SK 0y)	9.61 (6.75–12.71)	24.10 (17.54–42.86)
Poland Southeast		
2007 (PL 03)	6.29 (3.49–12.48)	15.86 (8.96–86.93)
2008 (PL 03)	*	
Portugal South		
2008 (P 01) ²	3.66 (2.46–4.78)	11.90 (8.80–20.04)
Romania East		
2008 (RO 02)	14.29 (10.56–19.19)	38.18 (27.16–65.65)
2009 (RO 03, RO 08, RO 09)	9.75 (7.07–13.03)	28.38 (20.20–48.72)
Romania West		
2008 (RO 05, RO 06, RO 07)	8.53 (7.06–10.19)	22.07 (17.67–30.12)
2009 (RO 10)	7.73 (3.50–22.49)	21.11 (10.36–422.61)
Spain Central		
2009 (ES 07, ES 08)	3.09 (2.03–4.33)	11.98 (8.12–22.31)

Table 4 continued

Population	MIC ₅₀ (95% CI) ^a	MIC ₉₀ (95% CI) ^a
Spain Northeast		
2008 (ES 04, ES 06)	7.03 (4.89–10.03)	23.91 (15.76–46.84)
2009 (ES 05, ES 06, ES 09)	6.40 (5.32–7.75)	13.68 (10.77–20.02)
Spain Southwest		
2008 (ES 01, ES 02, ES 03)	3.39 (2.94–3.89)	6.90 (5.79–8.89)

¹ for single values see table 3, ² pooled bioassay of three sites

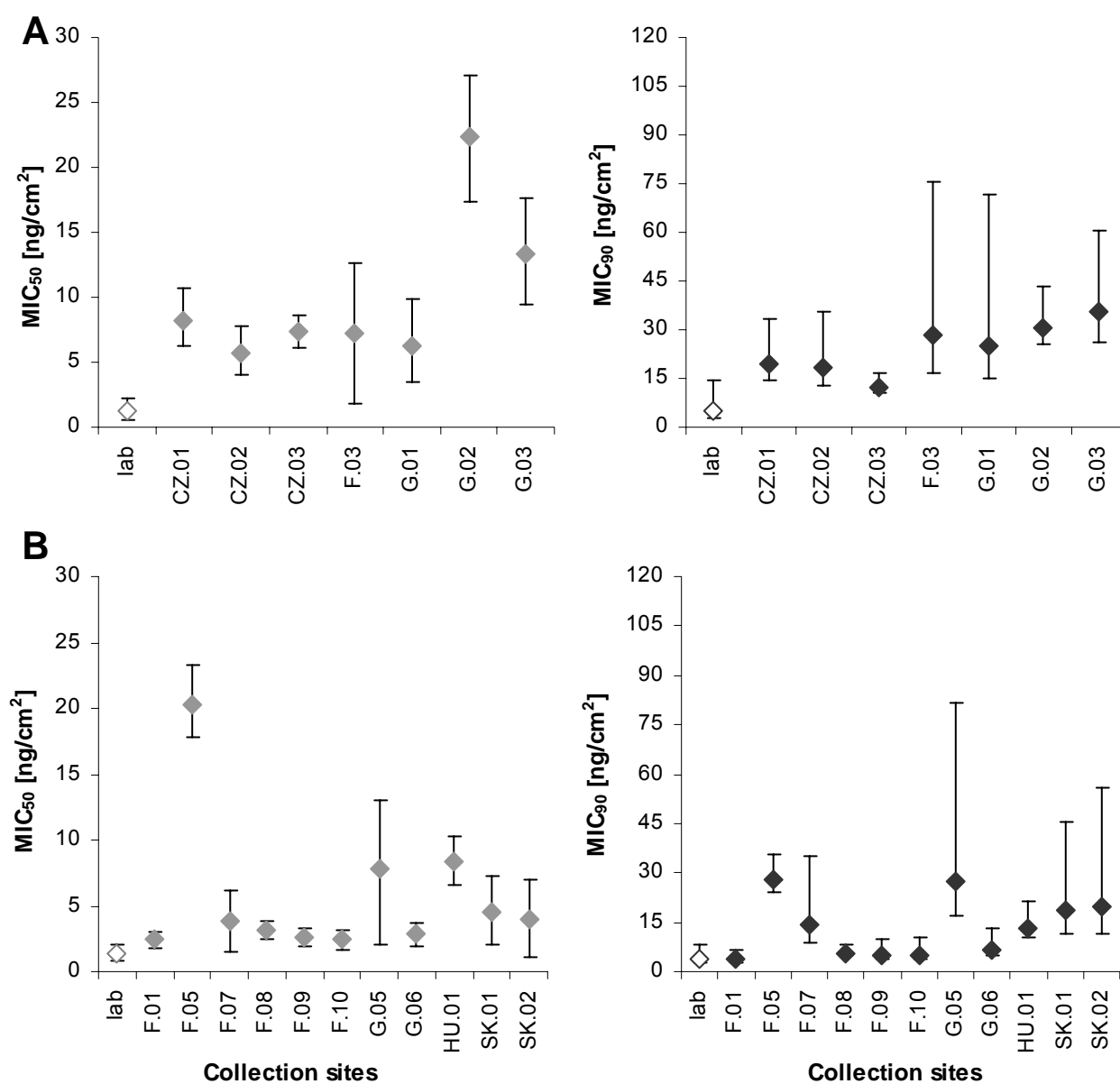


Figure 7: MIC₅₀ (left hand site) and MIC₉₀ (right hand site) ± 95 % CI of ECB neonates exposed to Cry1Ab collected in 2005 (A) and 2006 (B), (Legend see Tab. 4; lab - reference strain)

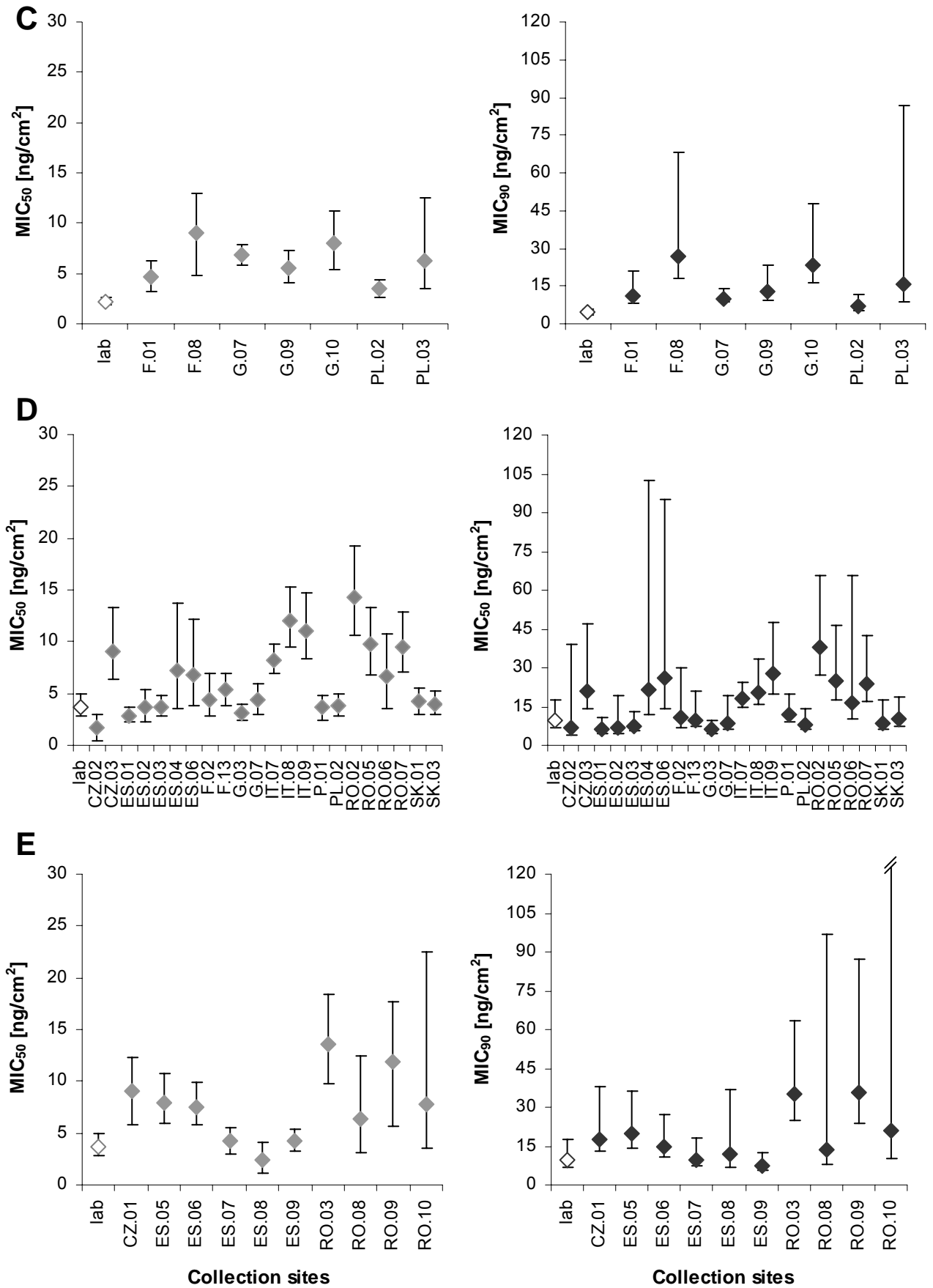


Figure 7: (continued) MIC₅₀ (left hand site) and MIC₉₀ (right hand site) ± 95 % CI of ECB neonates exposed to Cry1Ab in 2007 (C), 2008 (D), 2009 (E) (Legend see Tab. 4; lab - reference strain)

4. Conclusions

During 2005–2009, 15 populations with 98 samples (including replicates and assays without concentration response relationship) of ECB were analysed. Thus far, susceptibility to Cry1Ab have been assessed for one laboratory colony and populations collected in maize fields in Czech Republic, France, Germany, Italy, Hungary, Slovakia, Poland, Portugal, Romania, and Spain. ECB larvae were exposed to artificial diet treated with increasing Cry1Ab concentrations, and mortality and growth inhibition were evaluated after 7 days. Variation in Cry1Ab susceptibility of samples was up to 13.2-fold. A smaller variability was found for populations pooled according to geographic and climatic conditions (up to 6.6-fold). The results indicate that the observed population variation in susceptibility reflects natural variation in Bt susceptibility among ECB populations. Any evidence for a decrease of Cry1Ab susceptibility of populations during the monitoring duration from 2005–2009 could not be detected.

Further analyses have to be done to evaluate if the European populations of ECB are uniformly susceptible to Cry1Ab without any obvious genetic differentiation linked to geographical or other factors. In the future, other regional sources may be added to ensure that the monitoring program continues to represent the Cry1Ab maize market in Europe.

5. Acknowledgement

This report presents the results of research only and would not be possible without the kind help of all those who supplied insects: [REDACTED]

[REDACTED]. All those persons are key public sector entomologists or local industry contacts.

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7. Appendix (graphs show probit analysis results of assays conducted 2010)

Spain: Northeast Spain (ESne)

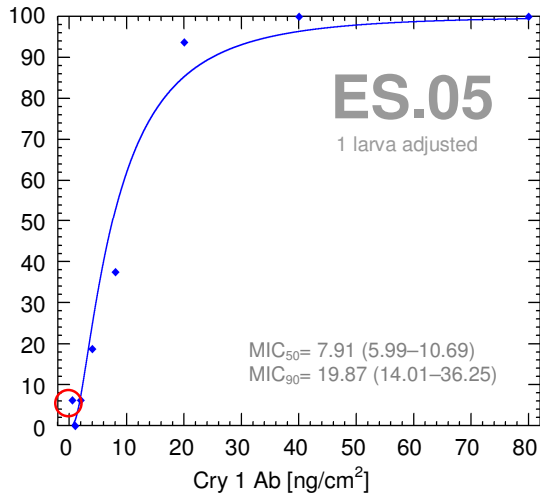


Fig. A1: Candasnos (ES.05), Spain

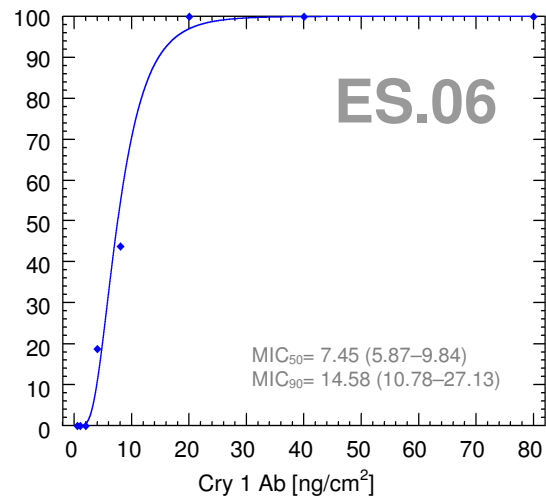


Fig. A2: Ejea de los Caballeros (ES.06), Spain

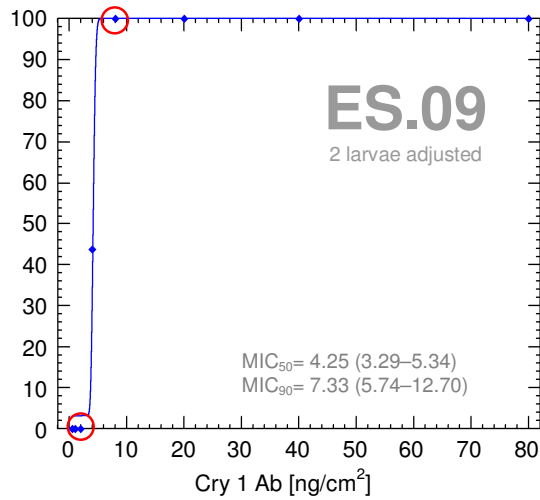


Fig. A3: Bujaraloz (ES.09), Spain

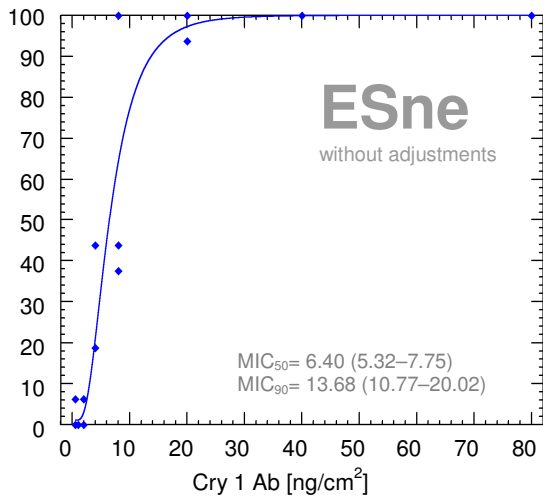
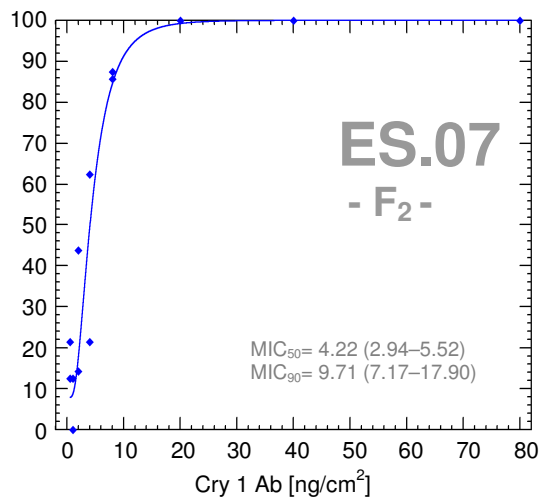
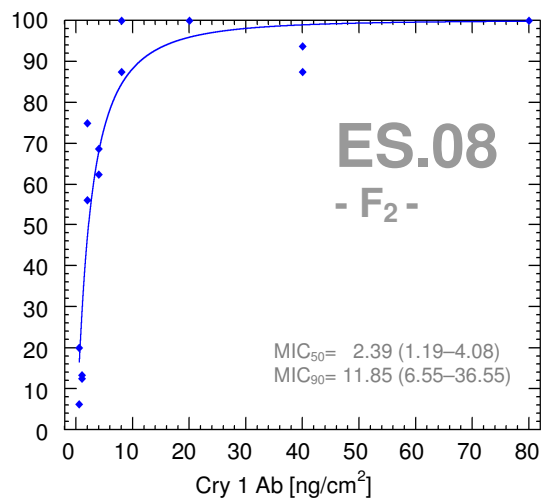
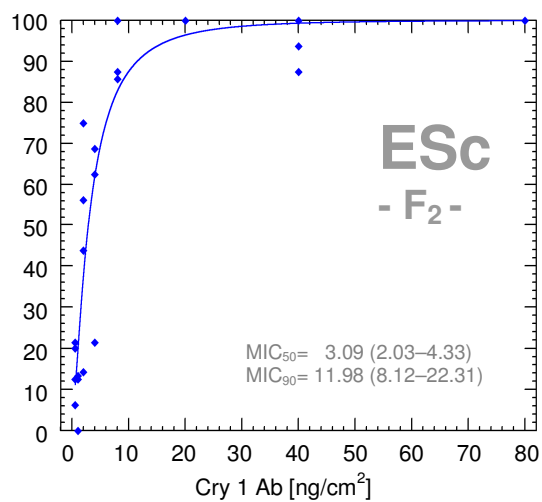


Fig. A4: Population (ES.05, ES.06, ES.09)
Spain

Spain: Central Spain (ESc)

Fig. A5: Aguas Nuevas (ES.07, F₂), SpainFig. A6: El Salobral (ES.08, F₂), SpainFig. A7: Population (ES.07 [F₂], ES.08 [F₂]), Spain

Romania: West Romania (ROw)

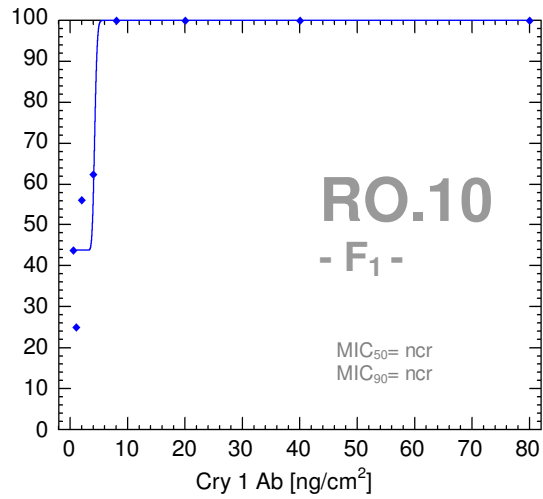
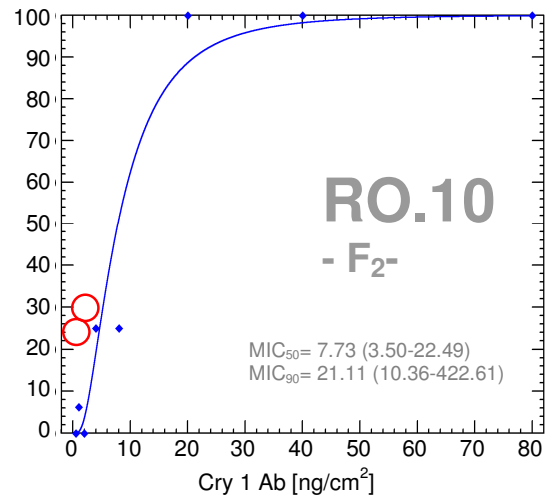
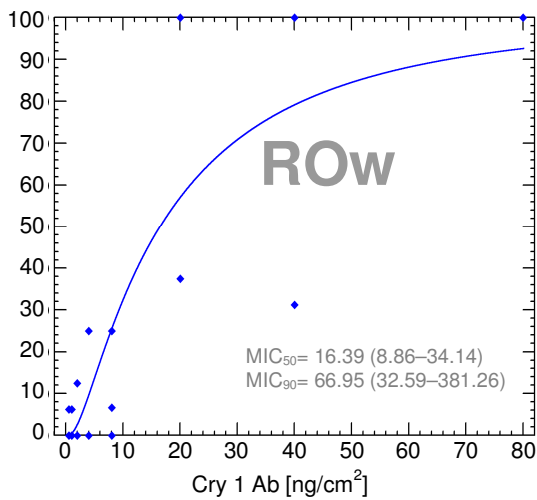
Fig. A08: Nadlac (RO.10, F₁), RomaniaFig. A09: Nadlac (RO.10, F_x), Romania

Fig. A10: (ROw), Romania

Romania: East Romania (ROe)

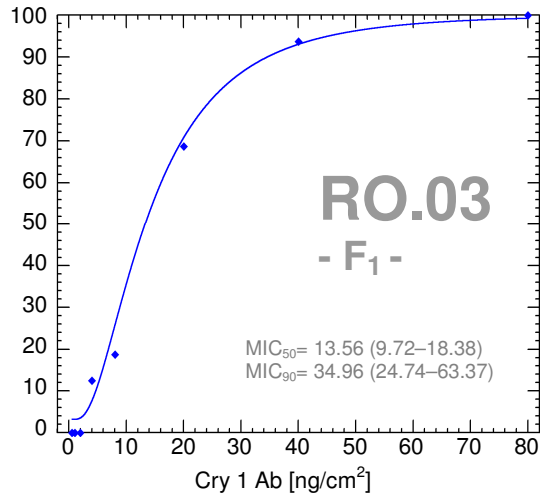


Fig. A11 Cascioarele (RO.03, F₁), Romania

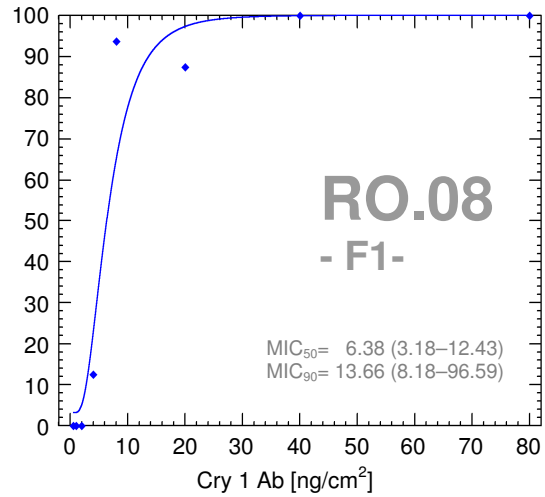


Fig. A12 Berceni (RO.08, F₁), Romania

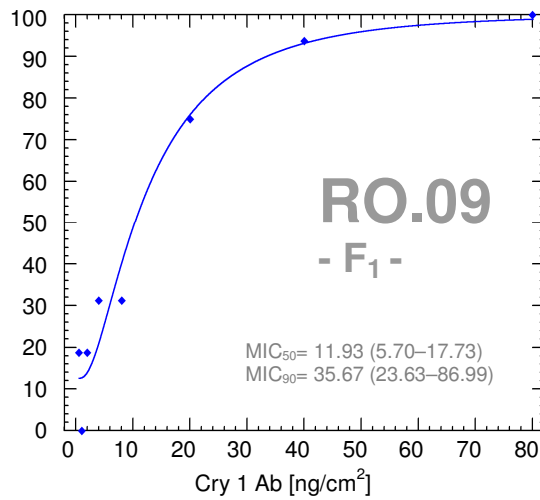


Fig. A13 Tudor Vladimirescu (RO.09, F₁), Romania

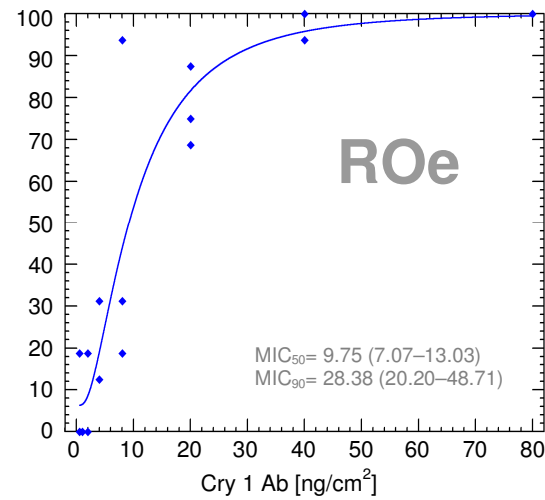


Fig. A14 (ROe), Romania